UNIVERSITY OF KENTUCKY
APPLICATION FOR CHANGE IN EXISTING COURSE: MAJOR & MINOR

1. Submitted by College of ______________________________ Department/Division offering course ______________________________ Date ______________________________

   Health Sciences   Clinical and Reproductive Sciences

2. Changes proposed:
   (a) Present prefix & number   CSC 624   Proposed prefix & number   CSC 624

   (b) Present Title   Gamete and Embryo Cryopreservation

   New Title   Cryopreservation of Reproductive Tissues

   (c) If course title is changed and exceeds 24 characters (Including spaces), include a sensible title (not to exceed 24 characters) for use on transcripts:

   Cryopreservation of Repro Tissues

   (d) Present credits:   1   Proposed credits:   2

   (e) Current lecture: laboratory ratio   1:1   Proposed:   1:1

   (f) Effective Date of Change: (Semester & Year)   Fall, 2008

3. To be Cross-listed as: ______________________________

   Prefix and Number

   Signature: Department Chair

4. Proposed change in Bulletin description:
   (a) Present description (including prerequisite(s)): Principles of cryopreservation will be covered; includes sessions on cryopreservation of human sperm and mouse embryos. Legal, ethical and policy issues associated with cryopreservation will be introduced. Prereq: CSC 620 and CSC 621.

   (b) New description: Principles of methods of cryopreservation and will be covered and procedures for freezing human oocytes, embryos, and ovarian and testicular tissues will be detailed. Legal, ethical and policy issues associated with cryopreservation will be introduced. Prereq: CSC 620 and CSC 621. Laboratory sessions will focus on freezing human spermatozoa and mouse gametes and embryos.

   (c) Prerequisite(s) for course as changed:   CSC 620 and CSC 621

5. What has prompted this proposal? Significant advances have been made in the cryopreservation of human reproductive tissues since the development of this course, including successful freezing of blastocysts, mature and immature oocytes and testicular and ovarian tissues. In order to remain current these topics must be included in the CSC 624 course. The change in credit hours is the results of both the added objectives and, based on experience, the time required to progress through the course.

6. If there are to be significant changes in the content or teaching objectives of this course, indicate changes:
   Objectives will be added to cover cryopreservation of: (1) blastocysts; (2) mature oocytes; (3)immature oocytes following maturation in the lab; (4) ovarian tissue; and (5) testicular tissue. Objectives addressing the policy, ethical and legal issues resulting from the freezing of these human oocytes and tissues also will be added.

7. What other departments could be affected by the proposed change? None

8. Is this course applicable to the requirements for at least one degree or certificate at the University of Kentucky?   X Yes   ☐ No
9. Will changing this course change the degree requirements in one or more programs? □ Yes □ No
   If yes, please attach an explanation of the change. (NOTE – If "yes," program change form must also be submitted.)

10. Is this course currently included in the University Studies Program? □ Yes □ No
   If yes, please attach correspondence indicating concurrence of the University Studies Committee.

11. If the course is 400G or 500 level, include syllabi or course statement showing differentiation for undergraduate and graduate students in assignments, grading criteria, and grading scales. □ Check here if 400G-500.

12. Is this a minor change? □ Yes □ No
   (NOTE: See the description on this form of what constitutes a minor change. Minor changes are sent directly from the Dean of the College to the Chair of the Senate Council. If the latter deems the change not to be minor, it will be sent to the appropriate Council for normal processing.)

13. Within the Department, who should be consulted for further information on the proposed course change?

   Name: Dr. Doris Baker
   Phone Extension: 323-1100 X80854
   Signatures of Approval:
   Date of Approval by Department Faculty 08/22/07
   Date of Approval by College Faculty 11/07/07
   Reported by Department Chair
   Reported by College Dean
   Reported by Undergraduate Council Chair
   Reported by Graduate Council Chair
   Reported by HCCC Chair
   Reported by Senate Council Office
   Reported by Senate Council Office

   *Date of Approval by Graduate Council 12/18/07
   *Date of Approval by Health Care Colleges Council (HCCC)
   *Date of Approval by Senate Council

   *Date of Approval by University Senate
   *If applicable, as provided by the Rules of the University Senate.

   *******

The Minor Change route for courses is provided as a mechanism to make changes in existing courses and is limited to one or more of the following:

a. change in number within the same hundred series;
b. editorial change in description which does not imply change in content or emphasis;
c. editorial change in title which does not imply change in content or emphasis;
d. change in prerequisite which does not imply change in content or emphasis;
e. cross-listing of courses under conditions set forth in item 3.0;
f. correction of typographical errors. [University Senate Rules, Section III - 3.1]

Rev 7/06
Although CSC 624 is a required course for the Master of Science in Clinical Sciences, reproductive laboratory science (RLS) track an increase of one credit hour does not affect degree requirements because students must complete a minimum of 30 credit hours to earn the degree. There is a range in credit hours depending on the student’s background and selected courses. For example, clinical laboratory science graduates do not take CSC 528 and hours for statistics, science and research courses vary (see table below). Currently the required courses in RLS are 19-23 hours and would increase to 20-22 credit hours. The remaining hours required for the degree will be fulfilled with statistics and science courses. Note also that students must complete 3 credit hours of RLS research and one hour in RLS seminar, but may elect to take additional hours in these courses.

### Masters of Science in Clinical Sciences - Reproductive Lab Science

<table>
<thead>
<tr>
<th>Statistics</th>
<th>Requirement may be fulfilled by taking:</th>
</tr>
</thead>
<tbody>
<tr>
<td>3-4 credit hours</td>
<td>• CSC 605 Epidemiology &amp; Biostatistics (3 credit hours) or</td>
</tr>
<tr>
<td></td>
<td>• CSC 606 Advanced Laboratory Statistics &amp; Administrative Analysis (3 credit hours) or</td>
</tr>
<tr>
<td></td>
<td>• STA 570 Basic Statistical Analysis (4 credit hours), or equivalent</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Physiology or Pathophysiology</th>
<th>Requirement may be fulfilled by taking:</th>
</tr>
</thead>
<tbody>
<tr>
<td>3-4 credit hours</td>
<td>• CSC 600 Pathophysiology (4 credit hours) or</td>
</tr>
<tr>
<td></td>
<td>• NURS 653 Pathophysiology (3 credit hours), or equivalent</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Science courses</th>
<th>To be selected from the following areas:</th>
</tr>
</thead>
<tbody>
<tr>
<td>3-6 credit hours (depending on background)</td>
<td>• Cell &amp; Molecular Biology</td>
</tr>
<tr>
<td></td>
<td>• Genetics</td>
</tr>
<tr>
<td></td>
<td>• Endocrinology</td>
</tr>
<tr>
<td></td>
<td>• Microbiology</td>
</tr>
<tr>
<td></td>
<td>• Other: Courses are selected in consultation with the student's advisory committee (3-6 credit hours minimum)</td>
</tr>
</tbody>
</table>

### Reproductive Laboratory Science Courses

<table>
<thead>
<tr>
<th>Course (semester hour credit)</th>
<th>Delivery Method *</th>
</tr>
</thead>
<tbody>
<tr>
<td>CSC 615 Reproductive Laboratory Science (1)</td>
<td>DL</td>
</tr>
<tr>
<td>CSC 616 Andrology (1)</td>
<td>DL</td>
</tr>
<tr>
<td>CSC 617 Reproductive Microbiology &amp; Immunology (1)</td>
<td>DL</td>
</tr>
<tr>
<td>CSC 528 Laboratory Techniques (2)</td>
<td>M, 2 weeks of 8 wk Summer Session</td>
</tr>
</tbody>
</table>

*Students not having an acceptable laboratory*
<table>
<thead>
<tr>
<th>Course Code</th>
<th>Course Name</th>
<th>Semester</th>
</tr>
</thead>
<tbody>
<tr>
<td>CSC 618</td>
<td>Labs in Andrology, Reproductive Microbiology &amp; Immunology (1)</td>
<td>M, 1 week of Fall Semester</td>
</tr>
<tr>
<td>CSC 621</td>
<td>Embryology &amp; ART (3)</td>
<td>Fall Semester</td>
</tr>
<tr>
<td>CSC 624</td>
<td>Gamete &amp; Embryo Cryopreservation (1)</td>
<td>Fall Semester</td>
</tr>
<tr>
<td>CSC 625</td>
<td>Mgt, Policy, Ethical &amp; Legal Issues in ART (2)</td>
<td>Fall Semester</td>
</tr>
<tr>
<td>CSC 626</td>
<td>Andrology Clinical Practicum (2)</td>
<td>1 weeks TBD</td>
</tr>
<tr>
<td>CSC 627</td>
<td>ART Clinical Practicum (3)</td>
<td>2 weeks TBD</td>
</tr>
<tr>
<td>CSC 628</td>
<td>RLS Seminar (1-2)</td>
<td>TBD</td>
</tr>
<tr>
<td>CSC 630</td>
<td>RLS Research (3-5):</td>
<td></td>
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</tbody>
</table>

Students already employed in assisted reproductive technology may complete portions of the research project at their place of employment under the supervision of a qualified laboratory director. A UK faculty member will direct the research project and collaborate with the ART supervisor.

* DL = Distributive learning. DL courses include web-based instruction and testing, and self-paced, interactive CDs. DL courses are offered during the spring semesters to both Lexington campus students and distance learners.

* M = Modular. Modular courses are taught at the UK campus. Classes meet for approximately 8 hours/day.

* TBD = To Be Determined

**NOTE** = All Clinical practica will take place in Assisted reproduction laboratories in the U.S. under the supervision of Directors appointed as clinical faculty by the University of Kentucky.
CSC 624 Cryopreservation of Reproductive Tissues

PROFESSOR: Doris J. Baker, Ph.D., HCLD(ABB), MT(ASCP), CLS(NCA)
OFFICE: 126E CTW Building, 900 South Limestone Street, Lexington, KY 40536-0200
PHONE NUMBER: (859) 323-1100 ext. 80854
E-MAIL dbake0@email.uky.edu
ED. COORDINATOR: Kim Campbell M.S., MT(ASCP)
PHONE NUMBER: (859) 323-1100 ext. 80853
E-Mail kkcamp1@email.uky.edu

Course Description: Gamete & Embryo Cryopreservation; 2 credit hours; Lecture/Lab 1:1
credit hour. Prerequisites: CSC 620, and CSC 621 or consent of instructor. Lecture: Principles of cryopreservation will be covered. Protocols for freezing spermatozoa, testicular tissue, oocytes, ovarian tissue and embryos at various stages of development will be detailed. Problems associated with freezing particular tissue types also will be discussed. Laboratory: Students will cryopreserve human sperm and mouse embryos using both manual and automated methods. They will then thaw the frozen gametes and embryos and assess survival and survival rates using standard criteria. Legal, ethical and policy issues associated with cryopreservation of human reproductive tissue will be introduced, including potential for transmission of infectious disease and issues associated with the term of storage of embryos.

By the end of the course, the student will demonstrate that he/she will/can:

Lecture:
- Understand the principles of cryopreservation, including factors that affect survival of cryopreserved cells, effects of physical stress from freezing, and damage caused during the thawing process.
- Compare advantages and disadvantages of rapid and slow cooling methods.
- Describe vitrification and discuss advantages and drawbacks of using this method.
- Determine the warming rate for specific reproductive tissues.
- Describe the action of cryoprotectants and demonstrate knowledge of intracellular and extracellular cryoprotectants, including when each should be incorporated in the procedure.
- Freeze hamster and murine gametes and embryos using both manual and automated freezing methods for reproductive tissue.
- Outline appropriate record keeping for the cryobiology lab including patient consent, monitoring, and terms of storage.
- Describe at least one acceptable protocol for freezing semen, embryos, and blastocysts. Describe the methods for freezing human oocytes; explain the problems associated with freezing metaphase oocytes and discuss options using prophase I oocytes.
- Diagram the steps for freezing and storage of human embryos using at least two different
automated protocols.

- Describe methods for freezing human oocytes and discuss problems associated with cryopreserving oocytes.
- Describe protocol for freezing testicular tissue, including tissue preparation.
- Describe protocol for cryopreserving oocytes (mature and process for maturing and cryopreserving immatures) and ovarian tissue.
- Discuss oocyte maturation followed by vitrification.
- Describe microbiological and genetic testing for sperm donors, oocyte donors and embryo donors.
- Discuss problems associated with cryopreservation of human embryos.
- Use resources available, including the web, to identify case studies that demonstrate legal and ethical issues associated with cryopreservation of human reproductive tissue.
- Outline protocol for freezing reproductive tissue for patients undergoing radiation and/or chemotherapy; identify problems based on the type of malignancy and the age of the patient.
- Discuss current and impending regulation for reproductive tissue banking.
- Fully describe all FDA regulations and ASRM guidelines for cryopreservation and storage of human oocytes, cleavage stage embryos, spermatozoa, testicular tissues and ovarian tissues.
- Fully describe FDA regulations and ASRM guidelines for donating all types of reproductive tissues.

**Laboratory:**

- Freeze, store, thaw and perform post-thaw assessment of human sperm using a manual method and two different automated methods.
- Prepare cryoprotectants and appropriate media for cryopreservation of reproductive tissue, including
  - Human sperm
  - Mouse oocytes
  - Mouse embryos
  - Mouse blastocysts.
- Outline preparation of cryoprotectants and appropriate media for cryopreservation of human oocytes, early stage embryos and blastocysts.
- Perform vitrification of mouse oocytes and embryos.
- Freeze, store, thaw and perform post-thaw assessment for the following reproductive tissue:
  - Mouse embryos
  - Mouse blastocysts
  - Human spermatozoa
  - Set-up and perform quality assurance for cryobanking.

**Required Text:** None

**References:**

**Books:**


Safe Cryopreservation of Gametes and Embryos. Edited by Eileen McLaughlin, Allan Pacey and Thomas Elliott. World Wide Conferences on Reproductive Biology. Ladybrook


PUBLIC BILLS AND LAWS


GUIDELINES, WORKSHOP SUMMARIES, ACCREDITATION PROGRAMS


Guidelines and newsletters as applicable:

1. American Society for Reproductive Medicine Newsletter
2. The American Fertility Society Guidelines for Practice
3. ASRM Net News
4. Androlog
5. EmbryoMail
6. Kaiser Newsletter
7. Alpha International
8. FertiNet

Journals:

American Journal of Obstetrics & Gynecology
American Journal of Reproductive Endocrinology
Andrologia
Biology of Reproduction
Clinical Obstetrics & Gynecology
Contraception
Development
Developmental Biology
Endocrinology Reviews
Fertility & Sterility
Gamete Research
Human Reproduction
International Journal of Developmental Biology
Journal of the American Medical Association
Journal of Andrology
Journal of Assisted Reproduction
Journal of Reproduction & Fertility
Journal of Assisted Reproduction & Genetics
Journal of Clinical Endocrinology & Metabolism
Journal of In Vitro Fertilization & Embryo Transfer
Journal of Microbiology
Journal of Reproduction and Fertility
Grading: CSC 624
Exam I  40%
Exam II  40%
Lab reports  20%

Grading Scale:
90-100%  = A
80-89% = B
70-79% = C
below 70% = E

Course Policies:
- Students are expected to review materials received prior to the commencement of CSC 624.
  - Laboratory reports are to be completed by the student without assistance.
  - Class attendance is expected for all sessions. Please notify the professor directly if you find it necessary to miss a session. If a student is absent for more than 20% of the lectures/labs, s/he will be dropped from the course.
  - Please see the professor during the first two days of class if you have any conflicts in scheduling due to religious observances.

University Policies:
Excused absences: Acceptable reasons for excused absences are listed in Student’s rights and Responsibilities, Section 5.2.4.2. Briefly, these include serious illness, illness or death of someone in the student’s immediate family, University sponsored trips, major religious holidays, and other circumstances the instructor finds reasonable. NOTE: If you intend to be absent to observe a major religious holiday, you must notify Dr. Stewart in writing by Jan 29.

When there is an excused absence, you will be given the opportunity to make up missed work and/or exams. It is the student’s responsibility to inform the instructor of the absence, preferably in advance, but no later than one week after the absence.

Senate Rule 5.2.4.2 states that faculty have the right to request “appropriate verification” when students claim an excused absence because of illness or death in the immediate family. The University Health Services (UHS) will no longer give excuses for absences from class due to illness or injury. UHS forms can be date stamped to show that students went to the trouble of going to University Health Service, but it does not mean that students actually saw a physician or a nurse. If the faculty member wants further verification that a student kept an appointment with University Health Services (especially when there have been multiple or prolonged absences from class), the student will need to sign a release of information form (available from UHS) that will give permission for the staff to talk with the faculty member.

Inclement weather: In case of inclement weather or emergencies, class will be held unless the University administration cancels classes. For University closing of classes and offices, call the UK Infoline at 257-5684 or check UKTV Cable Channel 16. Students should use their judgment about coming to class.
  - Severe Weather: UK Policy/Information:
It is the policy of the University of Kentucky to keep all offices open and classes meeting as scheduled except under extraordinary conditions.

If severe weather should result in changes to the university schedule, the university will follow specific procedures about when those decisions are made and how they will be announced. Details of those procedures are available at [http://www.uky.edu/PR/News/severe_weather.htm](http://www.uky.edu/PR/News/severe_weather.htm).

All faculty, staff and students should note that announcements regarding the cancellation of classes and closure of offices, or a delayed opening will normally be made by 6 a.m. through the local news media. The most up-to-date and complete information will be available from the UK Infoline at 257-5684, UK TV Cable Channel 16, or the UK Web site at [http://www.uky.edu](http://www.uky.edu).

**Cheating and plagiarism:** Descriptions of what constitutes cheating and plagiarism are found in *Student Rights and Responsibilities, Sections 6.3.1 and 6.3.2*. Be aware that the minimum consequence for either offense is an “E” in the course. Suspension and dismissal from the University are also options.

**Classroom and Learning Accommodations:** Instructors will make reasonable accommodations for physical and/or learning disabilities that could inhibit student academic success. The Disability Resource Center certifies the need for and specifies the particular type of such accommodations on a student-by-student basis. Students seeking accommodations must submit this certification to the faculty. Contact the Center staff at 257-2754.

**Writing Skills:** Helping promote scholarship is more than simply teaching the subject matter -- all students need to improve and refine their skills in verbal and written expression. Regardless of discipline, faculty has the right -and the obligation- to expect students to use English properly in all aspects of the course (S.R.5.2.4.3). Instructors may ask students to rewrite papers, make writing style one of the grading criteria, and report a seriously deficient student to his/her college for remedial work.
### Example Calendar – Modular Course

<table>
<thead>
<tr>
<th>Lectures – Drs. Baker and Witmyer</th>
<th>Lab Reports due-dates TBD</th>
</tr>
</thead>
<tbody>
<tr>
<td>• Embryo assessment and handling</td>
<td></td>
</tr>
<tr>
<td>• Cryopreservation of Day 2 and Day 3 Embryos</td>
<td></td>
</tr>
</tbody>
</table>

| Lab – Drs. Baker and Witmyer, Ms. Campbell        |                          |
| • Complete freezing run                            |                          |
| • Embryo storage                                  |                          |
| • Embryo assessment                                |                          |
| • Prep for Tuesday’s lab                           |                          |
| • Discuss troubleshooting                         |                          |

**Exam I**

| Lecture – Drs Baker and Witmyer                  |                          |
| • Embryo assessment and handling                  |                          |
| • Cryopreservation of blastocysts                 |                          |

| Lab – Dr. Witmyer, Ms. Campbell                  |                          |
| • Complete freezing run                           |                          |
| • Thaw Day 2 and Day 3 embryos                    |                          |
| • Embryo culture                                  |                          |

| Lecture – Dr. Witmyer                            |                          |
| • Embryo assessment and handling                  |                          |
| • Cryopreservation of embryos in straws           |                          |

| Lab – Dr. Witmyer, Ms. Campbell                  |                          |
| • Complete freezing run                           |                          |
| • Thaw blastocysts and embryos from morning run   |                          |

| Lecture - Drs. Angle and Baker                   |                          |
| ➢ Vitrification                                  |                          |

| Lab – Dr. Angle and Ms. Campbell                 |                          |

| Lecture: Dr. Baker                              |                          |
| ➢ Freezing testicular and ovarian tissues        |                          |
| ➢ Cryopreservation of immature oocytes           |                          |

| Lecture: Dr. Baker                              |                          |
| ➢ FDA regulations and ASRM guidelines for cryopreservation of reproductive tissues. |                          |

**Exam II**